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Estimation of Human Toxicity From Animal Inhalation Toxicity Data:

1. Minute Volume-Body Weight Relationships Between Animals And Man

BY:

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19980129 033

October 1997

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# ESTIMATION OF HUMAN TOXICITY FROM ANIMAL INHALATION TOXICITY DATA: 1. MINUTE VOLUME - BODY WEIGHT RELATIONSHIPS BETWEEN ANIMALS AND MAN.

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### **ACKNOWLEDGEMENTS**

The authors would like to thank the members of the Information Services Group at DRES for their assistance with literature searches and the extended task of acquiring the scientific reference material required for these reviews. The assistance of Dr. O. Moss of Chemical Institute of Inhalation Toxicology, Research Triangle Park, NC, Dr. H. Snipes of Inhalation Toxicology Research Institute, Albuquerque, NM, Dr. S.E. Reutter of ERDEC, APG, MD and the staff of DSIS, Ottawa and the librarians at University of Calgary Medical Library and the Agriculture Canada Library at Lethbridge Research Station, Agriculture Canada, Lethbridge.

### **ABSTRACT**

The relationship between body weight (BW) and respiratory minute volume ( $V_m$ ) was reviewed by collecting a data base from the literature of minute volume rates that encompassed species from mice at 12 g body weight to horses and a giraffe at  $\approx 500$  kg body weight. The data were separated into anesthetized and non-anesthetized groups and juvenile animals were removed from the non-anesthetized group. The final data set of non-anesthetized animals contained 134 studies representing 2304 animals and 18 species. The data show a power-law (allometric) relationship between the minute volume and body weight. The scaling or allometric parameters in this power-law have been estimated using a linear regression of the logarithms of the minute volume against body weight. The resulting allometric equations were;

$$Log_{10}V_{m} = -0.286 + 0.802 Log_{10}BW$$
 or  $V_{m} = 0.518 BW^{0.802}$ 

From these equations a corresponding set of minute volumes were obtained for various body weights of humans eg. 15.6 L/min for a 70 kg human. The results of the analyses were compared to similar studies in the literature. The relationship is recommended for military uses because it is derived from non-anesthetized, young adult mammals which are expected to mimic the soldier.

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Table I

Comparison of minute volume/body weight data from various literature sources

Species/ Minute volume/Body weight ratio ( L.min <sup>-1</sup> .kg <sup>-1</sup> )							
Source	C&W [1]	Guy	ton [2]	Snipes [3]	EPA [4]	This review	
Anesthetic	yes :	yes	no	no	mix	no	
Mouse	0.66	1.15	1.239	1.3	1.152	1.491	
Hamster	-	0.500	0.665	-	0.731	0.474	
Rat	0.64	0.688	0.638	0.80	0.615	0.881	
Guinea pig	0.19	0.323	0.334	0.66	0.475	0.616	
Rabbit	0.26	0.387	-	-	0.332	0.413	
Cat	0.26	-	-	-	0.335	0.257	
Dog	0.25	-	-	0.36	0.258	0.368	
Monkey	0.29	-	0.322	0.29	0.257	0.374	
Baboon	-	-	-	-	-	0.365	
Goat	-	-	-	-	0.235	0.263	
Sheep	-	-	-	-	0.298	0.289	
Pig	-	-	-	-	0.403	0.138	
Donkey	-	-	-	-	0.159	0.163	
Pony	-	-	-	-	0.208	0.183	
Mule	-	-	-	-	0.248	0.123	
Horse	-	-	-	-	0.118	0.131	
Cow	-	-	-	-	0.253	0.201	
Man (70 kg u	nless notec	l)					
- quoted	0.0914	0.127ª	0.128	0.09	0.286		
- calculated <sup>b</sup>	0.146	0.142	0.127	0.18	0.164	0.223	
Breathing rate	es for Man	` '					
- quoted	6.4	8.7 <sup>b</sup>	8.7	20	20	-	
- calculated <sup>b</sup>	10.2	10.	8.9	12.4	10.6	15.6	

EPA values were derived either from the allometric equations given for combined species or as the average value of data presented for that species [4].

<sup>&</sup>lt;sup>a</sup> Value for 68.5 kg man.

<sup>&</sup>lt;sup>b</sup> Values calculated from the matching allometric equations (vide infra).

### **EXECUTIVE SUMMARY**

Bide, R.W., Armour, S.J. and Yee, E., "Estimation of human toxicity from animal inhalation toxicity data: 1. A review and update of the supporting data required", SR 673, Defence Research Establishment SUFFIELD, October, 1997. UNCLASSIFIED

### **BACKGROUND**

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In order to estimate human inhalation toxicity from animal inhalation toxicity data there must be an acceptable formula that describes the relationship between animal and human body sizes and respiration rates<sup>1</sup>. Of necessity, this relationship must be determined empirically from experimental information using a variety of species both smaller and larger than humans. Once the relationship is established, the relationship may be used, in conjunction with toxicity data, to estimate human toxicity, first, from data with small laboratory species and then with increasing confidence as data with larger species are added to the data base.

Because of variation between animals, between techniques and technologies, between anesthetized and non-anesthetized animals and between handling practices in the laboratories, the respiratory data collected over the last century show considerable variability. Fortunately, there are sufficient studies to provide large statistical numbers which tend to reduce the effects of biological, age and sex variability. Furthermore, the experimenters recognized the effects of anesthetics and deliberately collected a great deal of information for non-anesthetized animals. Despite this, some of the most recent reviews of the data have ignored anesthetic use with the result that the collected data and interspecies relationships have greater variability than necessary. In addition, the most recent major review includes a large amount of data from very young animals which again increases the apparent variability in the statistical comparisons between species.

For this review, intended for military uses, data from the literature have been deliberately selected to match the soldier with regard to age and physical development. Anesthetized animal data are reported but not used. Only mammalian data was considered. Data from very young mammals and from mammals adapted to marine life were eliminated. To obtain the closest match of the human to the animal data, the minute volume for the human was taken directly from the calculated allometric rather than arbitrarily assigning a human value as has been done in other studies.

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The science of allometry is the study of the biology of scaling *ie.* the effects of body size on physiology, physiognomy and metabolism. For in depth treatment of the topic see [5, 6].

Table II

Recommended values of human minute volumes calculated for a range of body weights

Body weight (kg)	Minute volume (L/min)	Body weight (kg)	Minute volume (L/min)	Body weight (kg)	Minute volume (L/min)	
55	12.9	75	16.5	95	20.0	
60	13.8	80	17.4	100	20.8	
65	14.7	85	18.3	105	21.6	
70	15.6	90	29.1	110	22.5	

Minute volumes are calculated from the recommended allometric equation.

Table III

Calculated values for the "Standard" laboratory animals

Species	Body weight (kg)	Minute volume (L/min)	Minute volume Body weight ratio
Mouse	0.025	0.0269	1.0760
Rat	0.250	0.170	0.681
Guinea pig	0.50	0.297	0.594
Rabbit	3.0	1.249	0.416
Dog	10.	3.281	0.328
Ferret	1.1	0.559	0.508
Monkey	4.5	1.729	0.384

### **RESULTS**

The data encompasses 106 papers covering 24 species involving 241 experimental groups and data from 3443 animals. The data were split into anesthetized and non-anesthetized groups. The data for non-anesthetized animals selected represents 18 species, in 134 studies and 2304 land mammals. The animals range in size from mice at 12 g to horses and a giraffe at 480 - 550 kg. Marine adapted mammals were excluded because of the adaptation to underwater life-style involving infrequent breathing and extended breath-holding. Since the ultimate aim was to predict the values in man from those in other animals, the data for man were not included but were calculated from the data. The resulting value for humans compared favourably to published values.

The data provide a linear relationship between the  $Log_{10}$  of body weight and the  $Log_{10}$  of minute volume<sup>2</sup>. The regression equation may be expressed as either;

$$Log_{10}V_m = -0.286 + 0.802 Log_{10}BW$$
 or  $V_m = 0.518 BW^{0.802}$ 

where units of minute volume (V<sub>m</sub>) and body weight (BW) are L/min and kg, respectively.

From these data, the corresponding value for human respiration for a 70 kg man was calculated to be 15.6 L/min and the corresponding values for a range of breathing rates are given in Table II. Comparing this to literature values for human respiration indicates that the "standard man" would be doing light exercise when breathing at this rate.

The relationship between dose and breathing rate is believed to be linear *ie* as the breathing rate increases, the minute volume increases and the amount of substance inhaled increases in direct proportion. Thus, a man breathing at 30 L/min would inhale 30/15.6 or 1.92 times as much substance as our standard man.

Many literature studies on toxicology do not contain body weight and/or minute volume data on the test subjects. In these cases, data for "standard" animals are used. Values calculated from the allometric relationship above for "standard" laboratory animals are provided in Table III for convenience.

### **CONCLUSION**

The relationship above is recommended for military use because it has been derived from a large data base representing unanesthetized, young adult mammals which should mimic the age and physiologic state of the soldier.

The minute volume is the average volume of air inhaled (or exhaled) in one minute. It varies with body size, excitement, work and tension. It is the product of tidal volume (one breath) and frequency (how many breaths/min).

### INTRODUCTION

The assessment of human hazards from chemical exposures, whether deliberate or accidental, requires a knowledge of the differential relationships of metabolism, physiology and respiration between man and other mammalian species<sup>1</sup>, particularly those species commonly used for laboratory and agricultural experimentation. The requirement is particularly important when inhalation exposures are to be assessed because the doses received are not directly defined (as in direct injection/feeding routes of chemical introduction) but rather inferred from atmospheric and respiration parameters. With most hazard assessments, there is little information concerning direct human exposures and the toxicity estimates must be made by extrapolating animal data to assess the human condition.

Comparative physiologists and, more recently, toxicologists have provided a wealth of information concerning the relative respiration rates, tidal volumes and other respiratory parameters and several comparative studies have been published [1, 2, 3, 4, 70, 118, 128] relating the body weight, body surface and alveolar surface areas to these parameters in mammals. As technology has improved, the methods of measuring these functions have progressed from procedures requiring anesthesia and surgical intervention [54] to direct measurements on intact animals in enclosed chambers [135] to more modern methods of direct instrumental measurements on restrained but otherwise unmolested animals [106, 118, 129, 133]. In the process, the effects of using no anesthetic [63, 119,], single and combinations of anesthetics [64, 78, 152] have been studied. In retrospect, it is clear that the animal data on respiratory function must be grouped such that the emotional and physiologic state of the animals is as similar as possible and the comparative human values must be matched to similar animal data.

In inhalation toxicology, the dose received by an animal is directly related to the amount of air inhaled [3, 7, 8]. The important inter-related respiratory measurements are the respiration rate, the tidal volume and the minute volume. Several authors have attempted to relate minute volumes to the body weight of animal species [1, 2, 3, 4, 7, 70]. Although the process is generally agreed, the data and resulting mathematical values are quite different (Table I). Although some convergence may be noted in the most recent compilations, these data sets have been promulgated by authors from the same institutes.

The study of the relationships between body size and the physiology and metabolism is the science of allometry. For in depth treatments of the science see References 5 and 6.

Upon investigation, it was apparent that the data in two cases [2, 4] were derived from a mixture of anesthetized and non-anesthetized animals and another [3] could not be readily traced to published data. For this report, a relatively large number of studies providing body weight/respiratory volume data have been collected, reviewed, tabulated and the data compared to previous attempts to related body weight and minute volume. Relationships between body weight and minute volume have been derived for anesthetized and non-anesthetized animals and simplified formulae provided to assist in human hazard assessment from animal toxicity data.

### **METHODS**

The data gleaned from the literature for physiologic condition, body weights, minute volumes and the numbers of animals used were tabulated and mean values for each species and the standard deviation of the data were calculated using FRAMEWORK IV. When the number of data in a group (N value) was less than 4, standard deviations were not reported. The data were divided into anesthetized and non-anesthetized groups. The values for body weight and minute volume were plotted for all the data in each group and for each species in each group and the results evaluated.

A linear regression was calculated for each major group using a log-log relationship between the minute volume and body weight.

The data analysis was conducted using S-PLUS Version 3.4 for UNIX running on a Hewlett-Packard model 715/50 workstation under HP-UX (Hewlett-Packard, 3000 Hanover St., Palo Alto, California). S-PLUS (Mathsoft, Inc., Seattle, Washington) provides an interactive and integrated computing environment consisting of a suite of software facilities for programming, graphics and statistical analysis. All the computations were preformed in double precision (64 bit precision of a 32 bit computer). The computed numbers are carried calculated with 14 decimal places.

### RESULTS

### EVALUATION OF DATA COLLECTED FROM THE LITERATURE

The full data collection encompasses 106 papers covering 24 species tabulating 241

experimental groups and data from 3443 animals. The animals range in size from mice at 12 g to horses and a giraffe at 480 - 550 kg. The data, summarized in Tables III and IV, were divided into non-anesthetized (Tables III, V) and anesthetized (Tables IV, VI) groups and treated separately. Although data for birds were available and have been used in other summaries [4], they were not included here. Data for man were also excluded because one aim of the study was to obtain theoretical data for man. Several papers shown in the tables are for marine adapted mammals [104, 105, 147]. These data were not included in the final data set because of the adaptation to underwater life-style involving periodic or infrequent breathing and extended breath-holding. Because the ultimate aim was to predict the values in adult man from those in other animals, the data from very young (baby) animals also were excluded. These excluded data are marked with single asterisks in the tables.

When the data for non-anesthetized animals of all species were plotted as Log<sub>10</sub> of minute volume vs Log<sub>10</sub> of body weight (Fig. 1) a straight line resulted (power-law curve) with the variability within acceptable limits. In this graph, the data for each species are identified. Analysis of the variability in this data indicated that several studies were indeed statistically separate from the main body of the data. These included the cat studies of Wang [152], the cotton rat data of Guyton [2] and the spiny anteater study by Bentley *et al* [62] which are marked with double asterisks in the data tables. With these data removed, the final data set contained data from 134 studies on 18 species and 2304 animals.

### Analysis of the data set

A model for the respiratory system [9] based on a geometrically self-similar (fractal) network of flexible cylindrical tubes shows that the minute ventilation,  $V_m$ , and body mass, **BW**, is characterized by an allometric scaling relation of the form

$$V_{\rm m} = k \; \rm BW^{3/4}$$

where k is a "universal" constant. The power of BW, ie. 3/4 or 0.75, is referred to as the scaling exponent or constant. When the collected data for non-anesthetized animals were fitted to this equation, the data showed an "anomalous" scaling for the minute ventilation against body mass (weight), with a scaling exponent close to but slightly higher than the theoretical value of 0.75.

The measured allometric scaling relationship for minute ventilation and body mass for the compiled data set is shown in Fig. 2. In particular,  $V_m$  displays a scaling range of

about four decades with respect to BW, over which the observed relationship is seen to increase linearly on a double logarithmic plot. We conclude that  $V_m$  follows a power-law distribution of the form

(2) 
$$\operatorname{Log}_{10}V_{\mathbf{m}} = b + a \operatorname{Log}_{10}BW$$
 or  $V_{\mathbf{m}} = k BW^{\mathbf{a}}$ 

where  $b = \text{Log}_{10}k$ . A power-law distribution indicates the lack of a characteristic scale. A linear regression of the data collected gives values<sup>2</sup> for the constants in Equation 2 of:  $b = -0.286 \pm 0.017$  and  $a = 0.802 \pm 0.011$ . This relationship explains 97.5% of the variation in the data. The least-squares fitted line is shown by the dotted line in Figure 2 and the solid lines delineate the 99% simultaneous confidence bands for the fitted line. In the form of Equation 2, the regression equation would be

(3) 
$$\text{Log}_{10}\mathbf{V_m} = -0.286 + 0.802 \text{ Log}_{10}\mathbf{BW}$$
 or  $\mathbf{V_m} = 0.518 \text{ BW}^{0.802}$   $r^2 = 0.975$ 

where the units of minute volume  $V_m$  and of body weight BW are L/min and kg, respectively.

A normal quantile plot of the residuals from the least-squares fitted line (viz., the difference between the measured minute ventilation rates and the minute volumes predicted by Equation 3: Fig. 3) shows that the residual distribution appears to be well approximated by a Gaussian distribution. Nevertheless, there does appear to be a small amount of skewness in the data at the extreme upper and lower tails of the residual distribution. Here, it is seen that the points in the upper and lower tails lie somewhat below and above the line, respectively, suggesting that the residual quantiles here are slightly smaller and larger, respectively, than what would have been expected for a Gaussian distribution. The influence of these points on the least-squares fit appears to be minimal. In particular, the use of a highly robust method for fitting a linear regression based on minimizing the sum of the q smallest squared residuals (ie, the least trimmed squares regression) gave essentially the same values for the parameters a and b [cf. Equation 2], namely  $b = -0.286 \pm 0.008$  and  $a = 0.798 \pm 0.007$ . Therefore, we consider the simple regression to be sufficient.

Interestingly, the observed allometric scaling relationship between  $V_m$  and BW yields a scaling exponent of 4/5 or 0.80, rather than the theoretical value of 3/4. The difference

Values are mean ± standard deviation

between these two values of scaling exponent is admittedly small, but this difference was statistically significant. Indeed, if a fitted function of the form of Equation 2 with a = 3/4 suffers from a lack of fit to the data, then the aspect of the underlying pattern not accounted for by the fit should be evident in the residuals. Figure 4 shows the residuals  $\epsilon = \text{Log}_{10} V_m - 0.75 \text{ Log}_{10} BW$  plotted against  $\text{Log}_{10} BW$ . Note that these residuals show a clear positive linear trend, implying that  $V_m$  varies with BW with a scaling exponent that is greater than the putative 3/4 provided by theory. The loess curve added shows the residual dependence of  $\epsilon$  on  $\text{Log}_{10} BW$ . However, a plot of the residuals  $\epsilon = \text{Log}_{10} V_m - 0.80 \text{ Log}_{10} BW$  versus  $\text{Log}_{10} BW$  (Fig. 5) does not appear to show any systematic trends. Indeed, fairing a loess curve through these residuals shows that the only variation appears to be a constant variation with  $\text{Log}_{10} BW$ ; in particular, the constant value indicated by the loess curve for the residual variation is about -0.29 which coincides (approximately or better) with the least-squares fitted value for  $b = -0.286 \pm 0.011$ .

A similar analysis was done with the data for anesthetized animals with a similar result. The resulting regression equations for the anesthetized animals were

(4) 
$$\text{Log}_{10}\mathbf{V_m} = -0.509 + 0.782 \text{ Log}_{10}\mathbf{BW}$$
 or  $\mathbf{V_m} = 0.310 \text{ BW}^{0.786}$   $r^2 = 0.927$ 

where the units of minute volume  $V_m$  and of body weight BW are L/min and kg, respectively.

There were concerns that the contribution from small laboratory animals, which are physically and physiologically removed from man, would be over-emphasized because the numbers of studies and animals involved. If there was an such an effect, re-calculating the data using values weighted with the animal numbers should change the result in one direction and re-calculating the data using the mean values for each species should have an opposite effect. When these two extra calculations were performed, the resulting allometric equations were not significantly different and there was no apparent trend in the results. In fact, the effects noted on the non-anesthetized and anesthetized data sets appeared to be random. No further consideration was given to this topic.

The minute volume for a 70 kg man was calculated from Equation 3 (non-anesthetized animals) to be 15.6 L/min. From Equation 4 (anesthetized animals) the human minute volume was calculated to be 8.6 L/min. The 15.6 L/min value is within the bounds of accepted values for men doing "mild activity" [10, 11, 12, 13, 14] and slightly lower than the values measured in soldiers carrying light loads while walking on treadmills [15].

### EVALUATION OF THE DATA PROVIDED BY PHALEN

There are three data sets quoted in a text by Phalen [7]. The first, taken from Crosfill and Widdicomb [1], is reproduced as part of Table I. The data are from anesthetized animals and generally show much lower breathing rates than the other studies. The second data table given in the book is adapted from a paper by Boyd and Mangos [70] which, in turn, was compiled from a number of papers describing of both non-anesthetized and anesthetized animals. The third table in Phalen's book is adapted from a paper by Mauderly [124] describing data from non-anesthetized animals. All of the original papers from these three studies have been obtained and the data is included in the data tables presented in this study. Comparison of the data in the three tables indicates the wide range of values for respiratory parameters that may be found in the literature. An allometric relationship was calculated for the Crosfill and Widdicomb [1] data to be

(5) 
$$\text{Log}_{10}V_{\text{m}} = -0.485 + 0.810 \text{ Log}_{10} \text{ BW}$$
 or  $V_{\text{m}} = 0.327 \text{ BW}^{0.810}$   $r^2 = 0.960$ 

From this relationship, the human respiratory minute volume was calculated to be 10.2 L/min for a 70 kg man. As the data were appropriately included in the data tables of this review, no further work was done with these data sets.

### **RE-EVALUATION OF GUYTON'S DATA**

The data used by Guyton [2; Table I] in his analysis were compiled and split into anesthetized and non-anesthetized groups and regression lines in the form of Equation 2 were calculated for each to be

Anesthetized

(6) 
$$\text{Log}_{10}\mathbf{V_m} = -0.386 + 0.750 \text{ Log}_{10}\mathbf{BW}$$
 or  $\mathbf{V_m} = 0.411 \text{ BW}^{0.750}$ 

Non-anesthetized

(7) 
$$\text{Log}_{10}V_{\mathbf{m}} = -0.458 + 0.763 \text{ Log}_{10}BW$$
 or  $V_{\mathbf{m}} = 0.348 \text{ BW}^{0.763}$ 

and Guyton's published equation, when adjusted to the units used in this study<sup>3</sup>, becomes

(8) 
$$\text{Log}_{10}\mathbf{V_m} = -0.412 + 0.750 \text{ Log}_{10}\mathbf{BW}$$
 or  $\mathbf{V_m} = 0.388 \text{ BW}^{0.750}$ 

From these equations, it is obvious that the regression lines of the Log-Log format are very close to parallel (the slopes (b) are similar) and slightly separated (different intercept (a) values). The original, all combined Guyton data set produces a regression line that falls between those for the anesthetized and non-anesthetized animals. The slope of the equation for non-anesthetized animals is slightly steeper (0.763 vs 0.75).

For a 70 kg human the three Equations 6, 7 & 8, may be evaluated to give minute volume values of 9950, 8909 and 9372 cm<sup>3</sup>/min or 10., 8.9 and 9.4 L/min for the minute volumes of men in similar physiologic states to the respective animal populations. These equations are very similar, differing slightly in slope and intercept with the result that the calculated human minute volumes are inverted - anesthetized less than non-anesthetized.

### **RE-EVALUATION OF SNIPES' DATA**

The data quoted by Snipes [3] was treated in a similar manner to obtain the equations

(9) 
$$\text{Log}_{10}V_{m} = -0.273 + 0.741 \text{ Log}_{10}BW$$
 or  $V_{m} = 0.533 \text{ BW}^{0.741}$ 

where the minute volume  $V_m$  is in L/min and body weight BW is in kg.

A value of 12.4 L/min was obtained from these equations for a 70 kg man in similar physiologic condition. This value is not the value of 20 L/min given by Snipes adapted from Schlesinger [13] and Snyder *et al.*[14] for a 70 kg man doing "light activity".

The units used by Guyton were cm<sup>3</sup>/min for the minute volume, V<sub>m</sub>, and grams for the body weight, BW.

### RE-EVALUATION OF THE EPA DATA SET

The United States Environmental Protection Agency (EPA) published recommended values [4] for animal growth, feeding, and respiration "... to be used only when the study under review does not report values for the biological variables required for the risk assessment...". The data base has 87 papers yielding 203 entries encompassing 2570 animals of 44 species. Of these, 8 human studies contribute 22 entries and 301 "animals". Six papers are of wildlife of 17 species and in 17 of the 27 entries the numbers of animals are not reported. Nine species are birds and reptiles. In the remainder, the common laboratory species predominate. The data also include a number of studies with very young animals. Many of the studies are more recent than those reviewed here and, in the case of laboratory animals, more detailed as the different strains of the laboratory species are separately identified. Twenty-two of the 87 references are common to the current review and the EPA document. Unfortunately, those common references include both anesthetized and nonanesthetized animals and a number of the titles of the other references indicate that anesthetics may have been employed.

The EPA documents [4, 16] also give allometric equations for many of the species represented in their data base. However, the correlation coefficients are poor in comparison to that of the total data base and calculations of the minute volumes for the "standard" body weights produce some values that are considerably different from the experimental data. The use of the allometric equations for the separate species would appear to result in greater errors than would result from application of the total data base equations and, therefore, the presence of these "recommended" equations is noted and the use of same is discouraged.

The summary tables in the EPA document list the units of body weight as kg and of respiratory volume as m<sup>3</sup>/day. The quoted equation for the relationship between daily respiration (minute volume) and body weight is

$$I = 0.66 \text{ BW}^{0.7579}$$

where I is the daily respiration in m<sup>3</sup>/day and BW is the body weight in kg. This equation becomes

(11) 
$$\text{Log}_{10}\mathbf{V_m} = -0.339 + 0.758 \text{ Log}_{10}\mathbf{BW}$$
 or  $\mathbf{V_m} = 0.458 \text{ BW}^{0.758}$   $r^2 = 0.96$ 

when the units of minute volume  $(V_m)$  are L/min and the body weights (BW) are in kg. We were unable to reproduce this regression from the published data base (data excluded by the EPA either included or excluded). Several attempts were made, without success, to reproduce the EPA equation with modified data including removing the human data, weighting the data by the animal numbers and working with the average values for each species. The following equations were calculated from the EPA data set

(12) 
$$\text{Log}_{10}V_{\text{m}} = -0.384 + 0.763 \text{ Log}_{10}BW$$
 or  $V_{\text{m}} = 0.413 \text{ BW}^{0.763}$   $r^2 = 0.960$ 

using minute volume (V<sub>m</sub>) in L/min, body weight (BW) in kg.

The minute volume values for a 70 kg human calculated from Equations 11 and 12 were 11.5 and 10.6 L/min, respectively.

### **DISCUSSION**

The object of the exercise was to compile the data needed to obtain an estimate of the inhalation toxicity to soldiers (man) from inhalation toxicity values determined for a variety of animal species. To achieve this, a relationship must be established between the respiratory physiology of the animals and that of man. Three relationships that have been used and documented in the past are described and compared above [2, 3, 4]. The earliest and most quoted is that of Guyton [2] who used a mixture of anesthetized and nonanesthetized animals. When these are separated, the data becomes scant. Although, in the body weight/minute volume relationships (correlation regression lines) the slopes are only slightly different (0.750 vs 0.763), the intercept values are different and the predicted minute volumes for a 70 kg man differ by about 10%. In the data reported by Snipes [3], there are fewer species, no large species and the data could not be readily confirmed in published studies or reports. The minute volume for man calculated from the data was not the same as that quoted with the animal data. However, the data are quite similar to those generated for the present study. The third data set, from the EPA, also contains a mixture of nonanesthetized and anesthetized animals, a number of studies with very young animals and, upon close examination, relatively few studies albeit that they are generally more recent than those quoted by Guyton, Snipes and in this review. Comparisons of the regression equations (allometric equations) within the species with those between species in the EPA

report clearly indicates that something is distorting the calculated results.

Although the allometric regression equation (Equation 8) of Guyton [2] could be reproduced, that of the EPA (Equation 6) could not. There are a number of plausible reasons for this. From the EPA reports [4, 16], it is neither clear what statistical approach nor what computational assistance (computer) were used to calculate their results and, further, nothing is said about mechanisms for use of incomplete data. There are many data in the EPA set that had missing animal numbers and there are birds and seals as well as reptiles (which were stated to have been disregarded). The discrepancies between our calculations and those reported by the EPA are a little large for round-off errors. Because the EPA reports do not give details concerning how the calculations were done in 1988, there is the possibility of computer concatenation of squared numbers in the statistical calculations if insufficient precision was used. Since we have not obtained all of the original documents referred to for the EPA study, there is also the possibility of typographical errors.

Theoretical considerations from allometric scaling models [9] indicate that the scaling exponent in the equations should be three-quarter power, in this case  $BW^{0.75}$ . The data of Guyton [2], both anesthetized and non-anesthetized, fit very well to the predicted theoretical value of 0.75 or 3/4. The slope of the published equation in the EPA study (Equation 10) is close to 0.75. However, the same data when recalculated (Equations 12) gave a higher value. The slope values calculated in this study are slightly above 0.80 or 4/5 and the statistics clearly show a significant difference between the 0.75 and 0.80 values (P < 0.01). The 0.80 value has been obtained by others [5, 6, 17, 18, 19] and "...small deviations from quarter-power scaling sometimes occur [20, 21]." [9]. The data obtained clearly indicate that no artificial attempt should be made to force the body weight/minute volume relationship to fit the 3/4 power theoretical value.

The inter-species relationship between minute volume and body weight calculated from the data assembled for this review is considered by the authors to be the best currently available for use for military purposes. First and foremost, the calculated relationships fit, very closely, to the experimental data available. The data used are all from non-anesthetized mammals. Data from the very young have been removed. The majority of the data are from young adult animals and so match the target human population. Although there is a preponderance of data from laboratory species, the species range from very small to very large animals (mouse to giraffe) with a reasonable number in the sizes close to man. Finally, the number of studies, species and animals included in the calculations are large enough that the addition of more data should only result in small refinements rather than major

alterations in the resulting body weight/minute volume relationship.

There remains a moot point; - would a calculation, weighted by animal numbers, provide an improved or better estimate of the relationship between body weight and minute volume? The statisticians will argue that the weighted values should be used. However, this adds importance, perhaps undue importance, to the data for small laboratory species because there are both more studies and more animals used per study. Biologically, therefore, the argument can be made to refrain from weighting the studies in an attempt to reduce the imbalance caused by large numbers of small laboratory animals. Indeed, in a recent US study [Reutter, S.A. & Wade, J.; personal communication], the average values for each species were used in an attempt to negate the imbalance in the numbers of large vs small animals. The current recommendation is to use the unweighted data from non-anesthetized animals and the corresponding theoretical human minute volumes to estimate human toxicities from animal toxicity data. Equations for anesthetized animals are presented so that they would be available for use with toxicologic studies in which anesthetics were employed.

The minute volume of resting healthy men apparently does not change with age in the adult [22]. Thus, for military purposes, there is no requirement to make corrections for age of the soldier. Since in general, men and women have similar minute volume to body weight ratios [14], there is no need to complicate matters with a separate set of respiratory values.

When toxicity, body weight and minute volume data are available, the data may be used directly to extrapolate the toxicity to a human estimate. However, in many cases the required respiratory and body weight measurements are not available for the subjects of toxicity studies and "standard" animal values must be applied. Examination of the data shown in Fig. 1 and Fig. 2 indicates that the minute volume and body weight vary in a similar manner both within and between species. The EPA study provides allometric equations for many of the species commonly used in toxicologic studies [4] albeit that the correlations reported for intra-species variation are not as good as that given for interspecies effects (probably because of the effect of very young animals on the relationships). From the mean and standard deviation values obtained in this study (Table IV), it is clear that the use of data for a "standard" animal will not create a large error if the test and "standard" animals are not too disparate. Indeed, the use of "standard" animal values allows "...assessments made by different individuals or groups at different times..." to be "... compared more clearly., allowing disagreements to focus on important scientific judgements

rather than more mundane and trivial differences in assumptions of body weight and other biological variables." [4]. "Standard" animal values, given in Table III, were calculated using the recommended allometric relationship (Equation 3). The agreement with the collected data means is generally good. The mouse has the poorest agreement for the common laboratory species.

Since the physiologic state of all of the animals in a comparison should be as close as possible, the authors consider the best approach to be to use of the breathing rate for the required size of human that is calculated from the data presented in this treatise eg. 15.6 L/min for a 70 kg human. The appropriate values for a range of human body weights are given in Table II. In the other studies quoted, there is no general agreement on the minute volume value used for the human. Guyton [2] used a value, 6 L/min, for "resting" man which is lower than the 9.4 L/min value calculated from the data presented. Snipes [3], quoting Snyder [14], recommended a value of 20 L/min for man doing light activity but the data provided for the animals calculates to a value of 12.4 L/min for man. The EPA uses a combined value, also based upon Snyder, which provides a daily value for a man resting 8 hr and doing mild activity for 16 hr. The EPA equation provide human value of 10.6 L/min which also differs from the reference 20 L/min value. The ICRP Reference Man [14] breathes 23 m³/day (15.9 L/min) during light activity (which is surprisingly close to the calculated value of 15.6 L/min from this study).

The allometric equations presented provide the relationship between body weight and minute volume for a fixed set of physiologic parameters. These equations cannot be used to calculate the toxicity for different breathing rates eg. for a 70 kg man breathing 30 L/min. The toxicity resulting from inhaling either an air dissolved chemical or an aerosol should be directly related to and vary with the volume of contaminated air inhaled which may be represented by the minute volume. Therefore, the calculation of toxicity values for breathing rates other than that indicated from the allometric relationship should be a simple inverse proportion (as the toxicity increases the LCt<sub>50</sub> or LD<sub>50</sub> decreases). For example, the LCt<sub>50</sub> value for 70 kg human breathing 30 L/min would be 15.6/30 or 0.52 times the LCt<sub>50</sub> value for a human breathing 15.6 L/min.

### **SUMMARY**

The literature values for respiration of many species of animals have been gathered, collated and reviewed. The data base selected from the data collected, representing unanesthetized, young adult, mammals cover 18 species tabulating 134 experimental groups and data from 2304 animals. The animals range in size from mice at 12 g to horses and a giraffe at 480 - 550 kg.

The relationship between body weight and respiratory minute volume was reinvestigated using a larger, more directed data base than those used in previous studies. An allometric regression equation relating body weight and minute volume was obtained;

$$Log_{10}V_{m} = -0.286 + 0.802 Log_{10}BW$$
 or  $V_{m} = 0.518 BW^{0.802} r^{2} = 0.975$ 

The corresponding value, obtained from this allometric equations, for a 70 kg human under similar physiologic conditions was estimated to be 15.6 L/min. A table of minute volumes for various human body weights is provided.

In the judgement of the authors, the allometric relationship and the data presented represent the best currently available for use in the estimation of human inhalation toxicity for awake and lightly active soldiers.

### REFERENCES

- 1. Crosfill, M.L. & Widdicomb, J.G., "Physical characteristics of the chest and lungs and the work of breathing in different mammalian species", J. Physiol., 158, pp. 1-14, 1961
- 2. Guyton, A.C., "Measurement of the respiratory volumes of laboratory animals", Amer. J. Physiol., 150, pp. 70-77, 1947
- 3. Snipes, M.B., "Species comparisons for pulmonary retention of inhaled particles", Chapter 7, <u>Concepts in inhalation toxicology</u>, First Edition, Editors McClelland, R.O. and Henderson, R.F., Hemisphere Publishing Corp. New York, 1988, pp. 196.
- 4. Blackburn, K., "Recommendations for and documentation of biological values for use in risk assessment", EPA/600/6-87/008, US Environment Protection Agency, February 1988.

- 5. Schmidt-Neilsen, K., <u>Scaling</u>; Why is animal size so important, Cambridge Univ. Press, Cambridge, 1984, pp. 99-101.
- 6. Calder, W.A., <u>Size function and life history</u>, Harvard Univ. Press, Cambridge, MA, 1984, pp. 98.
- 7. Phalen, R.F., <u>Inhalation studies: Foundations and techniques</u>, CRC Press, Boca Raton, Florida, 1984, Chapter 9, pp. 211-242
- 8. Salem, H., "Factors affecting toxicity", Chapter 2 of <u>Inhalation toxicology</u>; <u>Research methods</u>, <u>applications and evaluation</u>, Ed. by Salem, H., Marcel Dekker, Inc., New York, 1987
- 9. West, G.B., Brown, J.H. & Enquist, B.J., "A general model for the origin of allometric scaling laws in biology", Science, 276, pp. 122-126, 1997
- 10. Sargeant, A.J., Rouleau, M.Y., Sutton, J.R. & Jones, N.L., "Ventilation exercise studies with circulatory occlusion", J. Appl. Physiol., 50, pp. 718-723, 1981
- 11. Taylor, C., "Studies in execise physiology", Amer. J. Physiol., 135, pp. 27-41, 1941
- 12. <u>Respiration and Circulation</u>, Biological Handbooks, Fed. Amer. Soc. Expt. Biol. Med., Ed by Altman and Dittmer, pp. 78-81, 1971
- 13. Schlesinger, R.B., "Comparative deposition of inhaled aerosols in experimental animals and humans; A review", J. Toxicol. Environ. Health, 15, pp. 197-214, 1985
- 14. Snyder, W.S., Cook, M.J., Karhausen, L.R., Naset, E.S., Howells, G.P. & Tipton, I.H., "International Commission on Radiologic Protection No. 23; Report of the task group on reference man", Oxford, U.K., Pergamon Press, 1975
- 15. Myles, W.S. & Saunders, P.L., "The physiologic cost of carrying light and heavy loads", Eur. J. Appl. Physiol., 42, pp. 125-131, 1979.
- 16. "Methods for derivation of inhalation reference concentrations and application of inhalation dosimetry, EPA/600/8-90/066F, US Environment Protection Agency, October 1994, pp. 4-29.
- 17. McMahon, T.A. and Bonner, On size and life, Scientific American Library, New York, 1983.
- 18. Bonner, J.T., <u>The evolution of complexity by means of natural selection</u>, Princeton University Press, Princeton, NJ. 1983
- 19. Brown, J.H., Macroecology, Univ. of Chicago Press, Chicago, IL. 1995
- 20. Stahl, W.R., "Scaling respiratory variables in mammals", J. Appl. Physiol., 22, pp. 453-460, 1967

- 21. Bennett, P.M. and Harvey, P.H., "Active and resting metabolism in birds, allometry, phylogeny and ecology", J. Zool. (London), 213, pp. 327-364, 1987
- 22. Hemingway, A., Pocock, D. & Short, J.J., "Variation of basal respiration with age", J. Chron. Dis., 3, pp. 301-310, 1956
- 23. Bennett, P.M., "Structural and functional determinates of metabolic rate", Amer. Zool., 28, pp. 699-708, 1988.
- 24. Harvey, P.H. and Pagel, M.D., <u>The comparative method in evolutionary biology</u>, Oxford Univ. Press, Oxford, 1991.

### **BIBLIOGRAPHY FOR ANIMAL DATA**

- 51. Alarie, Y., Ulrich, C.E., Krumm, A.A., Haddock, R.H. & Jennings, H.J., "Mechanical properties of the lung in cynomologus monkeys", Arch. Envir. Health, 22, pp. 643-654, 1971.
- 52. Allenby, A.C., Anderson, D.W., Bailey, S.C., Black, R.M., Scawin, J.W. & Upshall, D.G., Personal communication.
- 53. Altman, P.L. & Dittmer, D.S., <u>Handbook of Biological Data</u>, Biological Handbooks, Federation of American Societies for Experimental Biology, Washington, DC., 1956, pp. 1581
- 54. Amdur, M.O. & Mead, J., "Mechanics of respiration in unanesthetized guinea pigs", Amer. J. Physiol., 192, pp. 364-368, 1958.
- 55. Amdur, M.O. & Mead, J., "The respiratory response of guinea pigs to sulfuric acid mist", Arch. Ind. Health, 18, pp. 407-414, 1958
- 56. Amoroso, E.C., Scott, P. & Williams, K.G., "The pattern of external respiration in the unanesthetized animal", Proc. Royal Soc. B, 159, pp. 325-347, 1964
- 57. Attinger, E.O. & Cahill, J.M., "Cardiopulmonary mechanics in anesthetized pigs and dogs", Amer. J. Physiol., 198, pp. 346-348, 1960
- 58. Barcroft, J., Boycott, A.E., Dunn, J.S. & Peters, R.A., "Observation on respiration and circulation in the goat", Quarterly J. Med., 13, pp. 35-45, 1919
- 59. Barrow, R.E., Vorwald, A.J. & Dormier, E., "The measurement of small animal respiratory volumes by capacitance respirometry", Amer. Indust. Hyg. Assoc. J., 32, pp. 593-598, 1971.
- 60. Bartlet, D. & Tenney, S.M., "Control of breathing in experimental anemia', Resp. Physiol., 10, pp. 384-395, 1970
- 61. Bartorelli, C. & Gerola, A., "Tidal volume, oxygen uptake, cardiac output, and body surface in the cat", Amer. J. Physiol., 205, pp. 588-590, 1963

- 62. Bentley, P.J., Herreid, C.F. & Schmidt-Neilsen, K., "Respiration of a monotreme, the echidna, *Tachyglossus aculeatus*", Amer. J. Physiol., 212, pp. 957-961, 1963
- 63. Berendt, R.F., "The effect of physical and chemical restraint on selected respiratory parameters of Macaca mulatta", Lab. Anim. Care, 18, pp. 391-394, 1968.
- 64. Besch, T.K., Ruble, D.L., Gibbs, P.H. & Pitt, M.L.M., "Steady state minute volume determination by body only plethysmography in juvenile Rhesus monkeys", Lab. Anim. Sci., 46, pp. 539-544, 1996.
- 65. Binns, R. & Clark, G.C., "Lung function in cynomologus monkeys and baboons", Folia primatologica, 1972; Quoted in [66].
- 66. Binns, R., Clark, G.C. & Simpson, C.R., "Lung function and blood gas characteristics in the rhesus monkey", Lab. Animals, 6, pp. 189-198, 1972
- 67. Bisgard, G.E., Ruiz, A.V., Grover, R.F. & Will, J.A., "Ventilatory control in the Hereford calf", J. Appl. Physiol. 35, pp. 220-226, 1973.
- 68. Blaxter, K.L., "The effect of iodinated casein on the metabolism of sheep", J. Agric. Sci., 38, pp. 207-215, 1948.
- 69. Blume, W.& Zollner, R., "Uber die Wirkung des pervitins auf die atmung der ratte und des meerschweinchens", Arch. Exptl. Pathol. Pharmakol., 202, pp. 21-43, 1943
- 70. Boyd, R.L & Mangos, J.A, "Pulmonary mechanics of the normal ferret", J. Apl. Physiol., Resp. Envir. Exercise Physiol., 50, pp. 799-804, 1981
- 71. Brody, S., Bioenergetics and Growth, Reinhold, NY, 1945.
- 72. Brooks, P.M., Richey, E.O. & Pickering, J.E., "Prompt pulmonary ventilation and oxygen consumption changes in rhesus monkeys associated with whole body gamma-irradiation", Rad. Res., 6, pp. 430-449, 1957
- 73. Bucher, K., "Vergleichende charakterisierung der lungenatmung einiger sauger", Helv. Physol. Acta, 7, pp. 470-475, 1949.
- 74. Chapin, J.L., "Ventilary response of the unrestrained and unanesthetized hamster to CO<sub>2</sub>", Amer. J. Physiol., 179, pp. 146-148, 1954
- 75. Chvaloval, M., Kuncova, M., Havrankova, J. & Palecek, F., "Regulation of respiration in experimental silicosis", Physiologica Bohemoslovaca, 23, pp. 539-547, 1974.
- 76. Colebatch, H.J.H. & Halmagyi, D.F.J., "Lung mechanics and resuscitation after fluid aspiration", J. Appl. Physiol., 16, pp. 684-696, 1961
- 77. Colebatch, H.J.H. & Halmagyi, D.F.J., "Effect of vagotomy and vagal stimulation on lung mechanics and circulation", J. Appl. Physiol., 18, pp. 881-887, 1963
- 78. Cuddihy, R.G., Brownstein, D.G., Raabe, O.G. & Kanapilly, G.M., "Respiratory tract deposition of inhaled polydisperse aerosols in beagle dogs", Aerosol Sci., 4, pp. 35-45, 1973
- 79. Damon, E.G., Mauderly, J.L. & Jones, R.K., "Early effects of intratracheal instillation of elastase on mortality, respiratory function and pulmonary morphometry of F-344 rats", Tox. Appl. Pharmacol., 64, pp. 465-475, 1982.

- 80. Davidson, J.T., Wasserman, K., Lillington, G.A. & Schmidt, R.W., "Effect of aging on respiratory mechanics and gas exchange in rabbits", J. Appl. Physiol., 21, pp. 837-842, 1966
- 81. Diamond, L. & O'Donnell, M., "Pulmonary mechanics in normal rats", J. Appl. Physiol. Resp. Envir. Exer. Physiol., 43, pp. 942-948, 1977
- 82. Dirnhuber, P., Green, D.M. & Stratton, J.A., "Effects produced by high concentrations of inhaled dibenz (b,f)-1,4-oxazepine (CR) in conscious monkeys", CDE TP 369, 1978. RESTRICTED.
- 83. Edginton, J.A.G., Grace, T.J., Price, P.N. & Swanston, D.W., CDE Porton Down, December 1989, Personal communication.
- 84. Edginton, J.A.G., Grace, T.J., Price, P.N., Rice, P. & Upshall, D.G., CDE Porton Down, May 1990. Personal communication.
- 85. Edginton, J.A.G., Marrs, T.C., Price, P.N. & Upshall, D.G., "The acute toxicity of T2 mycotoxin to the guinea pig by the inahalation and subcutaneous routes", TP 384, Chemical Defence Establishment Porton Down, December 1984. RESRTICTED
- 86. Ekberg, D.R., & Hance, H.E., "Respiration measurements in mice", J. Appl. Physiol., 15, pp. 321-323, 1960.
- 87. Gautier, H., "Pattern of breathing during hypoxia or hypercapnia of the awake or anesthetized cat", Resp. Physiol., 27, pp. 193-206, 1976
- 88. Gautier, H., "Chronic ventilatory effects of diazepam and barbiturates in conscious cats", Eur. J. Pharmacol., 100, pp. 335-341, 1984.
- 89. Gautier, H., "Effects of chronic administration of phenobarbital in low doses on control of breathing in the cat", Respir. Physiol., 64(3), pp. 314-316, 1986
- 90. Gautier, H. & Bonora, M., "Ventilatory response of intact cats to carbon monoxide hypoxia", J. Appl. Physiol., 55, pp. 1064-1071, 1983.
- 91. Gillespie, J.R., Tyler, W.S. & Eberly, V.E., "Pulmonary ventilation and resistance in emphysematous and control horses", J. Appl. Physiol., 21, pp. 416-422, 1966
- 92. Grace, T.J. & Hobbs, M.J., CDE Porton Down, 1996, Personal communication.
- 93. Guyton, A.C., "Measurement of the respiratory volumes of laboratory animals", Amer. J. Physiol., 150, pp. 70-77, 1947
- 94. Hales, J.R.S. & Findlay, J.D., "The oxygen cost of thermally-induced and CO2 induced hyperventilation in the ox", Resp. Physiol., 4, pp. 353-362, 1968.
- 95. Hall, W.C. & Brody, S., "Growth and development with special reference to domestic animals XXVI. The energy increment of standing over lying and the energy cost of getting up and lying down in growing ruminants...", Mo. Agr. Exp. Sta. Res. Bull., 180, pp. 5-31, 1933
- 96. Halmagyi, D.F.J. & Colebatch, H.J.H., "Ventilation and circulation after fluid aspiration", J. Appl. Physiol., 16, pp. 35-40, 1961
- 97. Hamlin, R.L. & Smith, C.R., "Characteristics of respiration in healthy dogs

- anesthetized with sodium pentobarbital", Amer. J. vet. Res., 28, pp. 173-178, 1967.
- 98. Hayden, R.O., "Cardiopulmonary measurements obtained by right and left cardiac catheterization of *Macaca mulatta*", AMRL-TDR-64-69, Biophysics Laboratory, Aerospace Medical Research Laboratories, Wright-Patterson Air Force Base, Ohio, USA, November 1964
- 99. Hemingway, A., "The panting response of normal unanesthetized dogs to measured dosages of diathermy heat", Amer. J. Physiol., 121, pp. 747-754, 1938
- 100. Hemingway, A. & Hemingway, C., "Respiration of sheep at thermoneutral temperature", Resp. Physiol., 1, pp. 33 37, 1966.
- 101. Hemingway, A., Robinson, R., Hemingway, C. & Wall, J., "Cutaneous and brain temperatures related to respiratory metabolism of the sheep", J. Appl. Physiol., 21, 1223-1227, 1966
- 102. Howell, L.L., "Effects of caffeine on ventilation during acute and chronic nicotine administration in rhesus monkeys", J. Pharm. Exp. Therap., 273, pp. 1085-1094, 1995
- 103. Howell, L.L. & Landrum, A.M., "Attenuation of hypoxia-induced increases in ventilation by adenosine antagonists in rhesus monkeys", Life Sci., 57, pp. 773-783, 1995
- 104. Irving, L., Solandt, O.M., Solandt, D.Y. & Fisher, K.C., "The respiratory metabolism of the seal and its adjustment to diving", J. Cell. Comp. Physiol., 7, pp. 137-151, 1936
- 105. Irving, L., Scholander, P.F. & Grinnell, S.W., "The respiration of the porpoise, *Tursiops truncatus*", J. Cell. Comp. Physiol., 17, pp. 145-168, 1941
- 106. Jacky, J.P., "A plethysmograph for long term measurements of ventilation in unrestrained animals", J. Appl. Physiol., 45, pp. 644-647, 1978
- 107. Jaspar, N., Mazzarelli, M., Tessier, C. & Milic-emili, J., "Effect of ketamine on control of breathing in cats", J. Appl. Physiol., 55, 851-859, 1983
- 108. Johanson, W.G. & Pierce, A.K., "A noninvasive technique for measurement of airway conductance in small animals", J. Appl. Physiol., 30, pp. 146-150, 1971
- 109. Karel, L. & Weston, R.E., "Respiration in Macaca mulatta (Rhesus monkey)", Proc. Soc. Exp. Biol. Med., 61, pp. 291-296, 1946
- 110. Katusky, E., "Vergleichende untersuchungen uber den elastischen lungenwiderstand", Helv. Physiol. Acta, 15, pp. 358-360, 1957
- 111. Kelly, J.F., Cugell, D.W., Patterson, R. and Harris, K.E., "Acute airway obstruction in rhesus monkeys induced by pharmacologic and immunologic stimuli", J. Lab. Clin. Med., 83, pp. 738-749, 1974
- 112. Kibler, H.H., "Environmental physiology and shelter engineering XLIII. Energy metabolism and cardiorespiratory activities in shorthorn, santa gertudis and brahmin heifers during growth at 50 and 80 deg F temperatures", Missouri Agricultural Station Research Bulletin 643, October 1957.
- 113. Kibler, H.H., "Environmental physiology and shelter engineering. LV. Energy

- metabolism and related thermoregulatory reactions in brown swiss, holstein and jersey calves during growth at 50F and 80F temperatures", Missouri Agricultural Research Station Research Bulletin 743, July 1960.
- 114. Kleiber, M., "Tidal air of laboratory animals", Science, 99, pp. 542, 1944
- 115. Lai, Y.-L., Tsuya, Y. & Hildebrandt, J., "Ventilatory responses to acute CO<sub>2</sub> exposure in the rat", J. Appl. Physiol., 45, pp. 611-618, 1978
- 116. Lees, M.H., Malinow, M.R. & Parer, J.T., "Cardiorespiratory function of the rhesus monkey during phencyclidine analgesia", in Delcourt, L.V. (Ed.), Termas de Medicina, Sao Paulo, Brazil-Homenagem, pp. 61-69, 1965.
- 117. Leong, K.J., Dowd, G.F. & MacFarland, H.N., "A new technique for tidal volume measurement in unanesthetized small animals", Can. J. Physiol. Pharmacol., 42, pp. 189-198, 1964
- 118. Likens S.A. & Mauderly, J.L., "Respiratory measurements in small laboratory mammals: A literature review", ITRI and Lovelace B&ERI, LF-68 & UF-48, Lovelace Biomedical & Environmental Institute, Albuquerque NM, July 1979.
- 119. Liu, C.T. & DeLauter, R.D., "Pulmonary function in conscious and anesthetized rhesus monkeys", Amer. J. vet. Res., 38, pp. 1843 -1848, 1977
- 120. Loosli, C.G., Robertson, O.H. & Puck, T.T., "The production of experimental influenza in mice by inhalation of atmospheres containing influenza virus dispersed as fine droplets", J. Infect. Diseases, 72, pp. 142-153, 1943
- 121. Maidment, M.P. & Upshall, D.G., "Retention of inhaled perfluoroisobutylene in the rat", J. Appl. Toxicol., 12, pp. 393-400, 1992
- 122. Mauderly, J.L., "Steady state carbon monoxide-diffusing capacity of unanesthetized beagle dogs", Amer. J. Vet. Res., 33, pp. 1485-1491, 1972.
- 123. Mauderly, J.L., "Influence of sex and age on the pulmonary function of the unanesthetized beagle dog", J. Gerontology, 29, pp. 282-289, 1974
- 124. Mauderly, J.L., "Evaluation of the Grade Pony as a pulmonary function model", Amer. J. Vet. Research, 35, pp. 1025-1029, 1974
- 125. Mauderly, J.L., "Effect of age on pulmonary structure and function of immature and adult animals and man", Fed. Proc., 38, pp. 173-177, 1979
- 126. Mauderly, J.L., "Ventilation, lung volumes and lung mechanics of young and old Syrian hamsters", Exper. Aging Res., 5, pp. 497-508, 1979.
- 127. Mauderly, J.L., "The effect of age on respiratory function of fischer-344 rats", Exper. Aging Res. 8, pp. 31-36, 1982
- 128. Mauderly, J.L. & Tesarek, J.E., "Nonrebreathing valve for respiratory measurements in unsedated small animals", J. Appl. Physiol., 38, pp. 369-371, 1975
- 129. Mauderly, J.L., Tesarek, J.E., Sifford, L.J. & Sifford, L. J., "Respiratory measurements of unsedated small laboratory mammals using nonrebreathing valves", Lab. Animal Sci., 29, pp. 323-329, 1979

- 130. Mazzarelli, M., Jaspar, N., Zin, W.A., Aranda, J.V. & Milic-Emili, J., "Dose effect of caffeine on control of breathing and respiratory response to CO<sub>2</sub> in cats", J. Appl. Physiol., 60, pp. 52-59, 1986
- 131. McCutcheon, F.H., "The mammalian breathing mechanism", J. Cell. Comp. Physiol., 37, pp. 447-476, 1951.
- 132. McMahon, T.A., Brain, J.D. & Lemott, S., "Species differences in aerosol deposition", Inhaled Particles IV, Pt 1, Ed. Walton, W.H., Pergamon Press, 1977, pp. 23-33.
- 133. Mead, J., "Control of respiratory frequency", J. Appl. Physiol., 15, pp. 325-336, 1960
- 134. Olson, E.B. & Dempsey, J.A., "Rat as a model for humanlike ventilatory adaptation to chronic hypoxia", J. Appl. Physiol.; Respirat. Environ. Exercise Physiol., 44, pp. 763-769, 1978
- 135. Palecek, F., "Measurement of ventilatory mechanics in the rat", J. Appl. Physiol., 27, pp. 149-155, 1969
- 136. Palecek, F. & Rochova-Mikulaskova, J., "Experimental emphysema in rats; an attempt to influence its production by gestagen administration", Physiol. Bohemoslov., 17, pp. 445-454, 1968
- 137. Palecek, F., Paleckova, M. & Aviado, D.M, "Emphysema in immature rats produced by tracheal constriction and papain", Arch. Environ. Health, 15, pp. 332-342, 1967
- 138. Pappenheimer, J.R., "Measurement of respiratory exchange and duration of slow-wave sleep in unanesthetized rats breathing gas mixtures", J. Physiol., 260, pp. 7P-9P, 1976
- 139. Pappenheimer, J.R., "Sleep and respiration of rats during hypoxia", J. Physiol., 266, pp. 101-207, 1977
- 140. Pappenheimer, J.R., Fencl, V., Heisey, S.R. & Held, D., "Role of cerebral fluids in control of respiration as studied in unanesthetized goats", Amer. J. Physol., 208, pp. 436-450, 1965
- 141. Patterson, J.L., Goetz, R.H., Doyle, J.T., Warren, J.V., Gauer, O.H., Detweiler, D.K., Said, S.I., Hoernicke, H., McGregor, M., Keen, E.N., Smith, M.H., Hardie, E.L., Reynolds, M., Flatt, W.P. & Waldo, D.R., "Cardiorespiratory dynamics in the ox and giraffe with comparative observations on man and other animals", Ann. N.Y. Acad. Sci., 127, pp. 393-413, 1965
- 142. Pickrell, J.A., Dubin, S.E. & Elliott, J.C., "Normal respiratory parameters of unanesthetized beagle dogs", Lab. Animal Sci., 21, pp. 677-679, 1971
- 143. Pitt, M.L.M., Personal communication, USAMRICD, January 1997.
- 144. Raabe, O.G., Yeh, H.C., Newton, G.J., Phalen, R.J. & Velazquez, D.J., "Deposition of inhaled monodisperse aerosols in small rodents", <u>Inhaled Particles, IV, pt 2</u>, Ed. Walton, W.H., Pergamon Press, 1977, pp. 3-20
- 145. Schutz, E., "Bestimmung der atemgrosse narkotisierten ratten", Arzneimittel-Forsch., 10, pp. 52-53, 1960

- 146. Siafakas, N.M., Bonora, M., Duron, B., Gautier, H. & Milic-emili, J., "Dose effect of pentobarbital sodium on control of breathing in cats", J. Appl. Physiol., 55(5), pp. 1582-1589, 1983
- 147. Spencer, M.P., Gornall, T.A. & Poulter, T.C., "Respiratory and cardiac activity of killer whales", J. Appl. Physiol., 22, pp. 974 -981, 1967
- 148. Stahl, W.R. & Malinow, M.R., "A survey of physiologic measurements in Macaca mulatta", Folia primat., 7, pp. 12-33, 1967
- 149. Swank, R.L. & Foley, J.M., "Respiratory, electroencephalo-graphic and blood gas changes in progressive barbiturate narcosis in dogs", J. Pharm. Exptl. Therap., 92, pp. 381-396, 1948
- 150. Thomas, D.P. & Fregin, G.F., "Cardiorespiratory and metabolic responses to treadmill exercise in the horse", J. Appl. Physiol., 50, pp. 864-868. 1981
- 151. Vinegar, A., Sinnett, E.E. & Leith, D.E., "Dynamic mechanisms determine functional capacity in mice, Mus musculus", J. Appl. Physiol., 46, pp. 867-871, 1979
- 152. Wang, S.C. & Nims, L.F., "The effect of various anesthetics and decerebration on the CO<sub>2</sub> stimulating action on respiration in cats", J. Pharm. Exper. Therap., 92, pp. 187-195, 1948
- 153. Watkins, B.E., Riegle, G.D. & Heisey, S.R., "Respiratory responses to ACTH and Dexamethazone in unanesthetized goats", Resp. Physiol., 17, pp. 127-133, 1973
- 154. Webber, C.L. & Peiss, C.N., "Pentobarbital-induced apneusis in intact, vagotomized and pneumotaxic-lesioned cats", Resp. Physiol., 38, pp. 37-57, 1979
- 155. Wolff, R.K., Kanapilly, G.M., DeNee, P.B. & McClellan, R.O., "Deposition of 0.1 μm chain aggregate aerosols in beagle dogs", J. Aerosol Sci., 12, pp. 119-129, 1981
- 156. Zin, W.A., Pengelly, L.D. & Milic-emili, J., "Active impedance of respiratory system in cats", J. Appl. Physiol., 53, pp. 149-157, 1982

Table IV

Average Body weights, minute volumes and MV/BWt for non-anesthetized animals

Species	Body Weight (kg)	Minute Volume (L)	Minute Volume B Wt	Species	Body Weight (kg)	Minute Volume (L)	Minute <u>Volume</u> B Wt
Cat	3.36 <sup>32,5</sup> ±0.480	0.845 ±0.061	0.257 ±0.052	Pony	167 <sup>3,3</sup> ±35	29.4 ±13.8	0.183 ±0.092
Cows	370 <sup>68,19</sup> ±147	71 ±33	0.189 ±0.047	Horse	465 <sup>27,4</sup> ±29	62 ±14	0.133 ±0.029
Donkey	120.1,1	19.5	0.16	Mule	210 <sup>2,2</sup>	25.9	0.12
Dog	13.0 <sup>739,21</sup> ±11.5	4.41 ±1.99	0.368 ±0.089	Sheep	52.6 <sup>46,11</sup> ±15.2	13.9 ±7.2	0.289 ±0.160
G pig	0.317 <sup>483,13</sup> ±0.129	0.179 ±0.039	0.616 ±0.161	Pig	197 <sup>2,2</sup> ±39	28 ±13	0.138 ±0.037
Goat	36.9 <sup>38,4</sup> ±12.3	9.6 ±2.8	0.263 ±0.052	Giraffe	544. <sup>2,1</sup>	74	0.136
Hamsters	0.111 <sup>97,4</sup> ±0.016	0.051 ±0.008	0.474 ±0.142	Rabbit	2.77 <sup>61,3</sup> ±0.63	1.453 ±0.193	0.413 ±0.787
Mouse	0.021 <sup>74,4</sup> ±0.007	0.031 ±0.014	1.491 ±0.366	Rat	0.259 <sup>314,21</sup> ±0.073	0.211 ±0.078	0.881 ±0.437
*Cotton rat	0.077 <sup>27,1</sup>	0.040	0.516	Baboon	4.42 <sup>4,1</sup>	1.61	0.365
*Spiny anteater	3.47 <sup>3.1</sup> ±0.177	0.527 ±0.251	0.150 0.065	Monkey	4.27 <sup>311,15</sup> ±2.69	1.40 ±0.42	0.374 ±0.125

Superscript values on the body weight data are (number of animals, number of studies)

<sup>\*</sup> Excluded from analysis, See text.

Table V

Average Body weight, minute volume and MV/BWt for anesthetized animals

Species	Body Weight (kg)		Minute <u>Volume</u> B Wt	Species	Body Weight (kg)	Minute Volume (L)	Minute <u>Volume</u> B Wt
Cat	3.06 <sup>94,12</sup> ±0.393	0.707 ±0.271	0.226 ±0.066	Giraffe	544. <sup>1,1</sup>	61.	0.111
Dog	11.6 <sup>135,11</sup> ±4.0	2.64 ±1.44	0.234 ±0.112	Rabbit	2.89 <sup>74,5</sup> ±1.12	0.67 ±0.14	0.251 ±0.085
Ferret	0.31418,1	0.195	0.621	Rat	0.289 <sup>397,22</sup> ±0.173	0.129 ±0.046	0.524 ±0.184
G pig	0.583 <sup>53,2</sup> ±0.151	0.142 ±0.017	0.256 ±0.095	Monkey,	3.75 <sup>127,23</sup> ±1.09	0.891 ±0.397	0.235 ±0.065
Hamsters	0.121 <sup>114,6</sup> ±0.015	0.043 ±0.016	0.372 ±0.170				
Mouse	0.025 <sup>46,6</sup> ±0.005	0.026 ±0.016	1.044 ±0.588				
Sheep	41.8 <sup>25,3</sup>	8.51 ±3.7	0.204 ±0.083				
Pig	<b>28</b> . 19,1	6.7	0.24				

Superscripts in the body weight column are (number of animals, number of experiments) involved in the mean values.

<sup>&</sup>lt;sup>a</sup> Body weight value for sheep is artificial, calculated from a minute volume provided as L/min/m² body surface using the formula given by the author for calculating surface area from body weight. The value for minute volume is the quoted value in L/min/m² and the body weight value is the corresponding weight in kg for one m².

Table VI

Comparisons of respiratory function and body weight Summary of data collected; non-anesthetized animals

Quiminai y	or data compoted,			JJ		
		Body	Number	Minute	Minute	
	Physiological	Weight	of	volume	<u>volume</u>	
Species	state and details	(kg)	animals	(Litres)	Body Wt	Reference
Monkey	chair, 30 min	2.80	60	1.487	0.531	63
Monkey	str. jacket, 30 min	2.80	60	1.587	0.567	63
Monkey	sedated, conscious	3.00	3	1.000	0.333	82
Monkey	sedated	3.50	16	1.200	0.343	83
Monkey	sedated	3.50	3	1.400		83
Monkey		2.68	6	0.863	0.322	93
Monkey, Rhesus	chair	10.70	3	2.000		103
Monkey, Rhesus	chair	10.90	6	2.000		103
Monkey, Rhesus	untrained, quieted	2.63	39	1.014	0.386	109
Monkey, Rhesus	untrained, quieted	3.49	4	0.757	0.217	109
Monkey, Rhesus	conscious, O <sub>2</sub>	4.05	4	1.100	0.272	119
Monkey, female, Rhesus	chair	3.33	8	1.820	0.547	66
Monkey, male, Rhesus	chair	3.48	8	1.441	0.414	66
Monkey, cynom	chair	4.25	10	2.039	0.480	65, 66
Monkey, cynom	chair	3.00	81	1.272	0.424	51
*Monkey, pygmy	Free, infant	0.68	1	0.400	0.588	106
Baboon	chair	4.42	4	1.612	0.365	65, 66
Cat	trained, free	3.00	4	0.804	0.268	60
Cat	trained, awake, not purring		6	0.840	0.247	88
Cat	trained, plethysmograph	2.80	11	0.950	0.339	87
Cat	trained	3.62	6	0.830	0.229	89
Cat	trained, plethysmograph	4.0	5	8.0	0.2	90
**Cat	trained, plethysmograph	2.23	4	0.475	0.213	152
**Cat	trained, plethysmograph	2.55	4	0.490	0.192	152
**Cat	trained, plethysmograph	2.58	4	0.373	0.145	152
**Cat	trained, plethysmograph	2.35	4	0.284	0.121	152
Cows,	<b>O</b> .	439.	4	78.4	0.179	141
*Calves, Ayrshire 8 mo	Standing, stanchion	147.	4	31.3	0.213	94
Cows, Br.Swiss,		250.	3	44.0	0.176	112, 156
Cows, Br.Swiss,	· ·	325.	3	50.2	0.154	112, 156
Cows, Brahmin	•	350.	3	37.9	0.108	112, 156
Cows, Brahmin,	•	250.	3	34.0	0.136	112, 156
Cows, Guernsey	-	435.	2	126.7	0.291	56
Cows, Guernsey,	Standing	410.	1	107.0	0.261	53, 95

<sup>\*</sup> Excluded as too young

<sup>\*\*</sup> Excluded as outlier, See text.

Table VI Continued; non-anesthetized animals

	Physiological state and details	Body Weight (kg)	Number of animals	volume	Minute volume B Wt	Reference
Cows, Hereford,	Standing	422.	1	109.0	0.258	53, 95
Cows, Hereford,	Standing	144.	1	49.0	0.340	53, 95
Cows, Holstein,	Standing	514.	1	114.0	0.222	53, 95
Cows, Holstein,	Standing	164.	1	61.0	0.372	53, 95
Cows, Holstein,	stanchion, raised 50F	250.	3	51.4	0.206	113
Cows, Holstein,	stanchion, raised 50F	350.	3	66.5	0.190	113
Cows, Jersey	stanchion	340	1	59.3	0.174	56
Cows, Jersey,	Standing	403.	1	92.0	0.228	53, 95
Cows, Jersey,	stanchion, raised 50F	225.	3	39.1	0.174	113
Cows, S.Gertrudis	stanchion, raised 80F	250.	3	49.7	0.199	113
Cows, S.Gertrudis	stanchion, raised 80F	350.	3	55.9	0.160	113
Cows, Shorthorn	stanchion raised 50F	250.	3	45.8	0.183	113
Cows, Shorthorn	stanchion raised 50F	350.	3	49.1	0.140	113
*Cows, Guernsey	stanchion, raised 50°F	110	3	5.6	0.051	112, 156
*Cows, Br.Swiss,	stanchion, raised 50F	100.	3	23.7	0.237	112, 156
*Cows, Brahmin,	stanchion, raised 80F	100.	3	19.1	0.191	112, 156
*Cows, Hereford, 4-6 wh		59.	8	13.1	0.222	67
*Cows, Holstein,	stanchion, raised 50F	100.	3	25.5	0.255	113
*Cows, Jersey,	stanchion, raised 50F	100.	3	21.9	0.219	113
*Cows, S.Gertrudis	stanchion, raised 80F	100.	3	27.4	0.274	113
*Cows, Shorthorn	stanchion, raised 50F	100.	3	35.4	0.354	113
Dog, Beagle		11.3		3.60	0.319	125
Dog, Beagle		11.2		3.90	0.348	125
Dog, Beagle 13 months,	5500 ft, Albuquerque NM	9.2	39	5.28	0.577	142
Dog, Beagle		10.5	20	3.14	0.299	155
Dog, Beagle	1 year	9.2	140	3.60	0.391	125
Dog, Beagle	5 year	11.2	48	3.90	0.348	125
Dog, Beagle	10 year	11.3	50	3.60	0.319	125
Dog, Beagle	restrained	9.6	49	4.47	0.465	122
Dog, Beagle, female	12-14 mo	8.4	50	3.60	0.429	123
Dog, beagle, female	3-4 yr	10.0	10	3.61	0.361	123
Dog, Beagle, female	8-10.5 yr	10.9	36	3.81	0.350	123
Dog, beagle, male	12-14 mo	10.0	50	3.65	0.365	123
Dog, beagle, male	3-4 yr	12.0	10	4.51	0.376	123
Dog, Beagle	trained, free	9.0	12	3.30	0.367	129
Dogs		9.0		3.70	0.411	118, 124
Dog	restrained plethysmograph	59	1	11.7	0.1981	56
Dog	restrained plethysmograph	13.5	1	6.6	0.489	56

<sup>\*</sup> Excluded as too young
\*\* Excluded as outlier, See text.

Table VI Continued; non-anesthetized animals

Species	Physiological state and details	Body Weight (kg)	Number of animals	volume	Minute volume B Wt	Reference
Dog *Dog, Beagle	restrained plethysmograph 0.25 year	15.6 3.1	5 10	3.41 1.80	0.219 0.581	99 125
G pig	free	0.466	61	0.156	0.334	93
G pig	trained, free	0.512	10	0.171	0.334	129
G pig	trained, free	0.194	8	0.162	0.835	133
G pig	trained, free	0.212	5	0.154	0.724	133
G pig	restrained	0.345	24	0.236	0.684	92
G pig	restrained	0.245	26	0.157	0.641	52
G pig	plethysmog, intrathoracic catheter	0.219	200	0.139	0.635	54
G pig	plethysmog, intrathoracic catheter	0.215	85	0.139	0.644	54
G pig	plethysmog, intrathoracic catheter	0.209	60	0.137	0.656	54
G pig	free	0.588	1	0.226	0.384	85
G pig	free	0.312	1	0.209	0.670	85
G pig	free	0.278	1	0.204	0.733	85
G pig	free	0.329	1	0.242	0.736	85
Goat	trained, masked	52.5	3	12.3	0.235	153
Goat	laterally recumbent, awake	23.7	21	5.7	0.242	58
Goat	stanchion	31.5	4	10.7	0.341	56
Goat	trained, masked	40.0	10	9.4	0.235	140
Hamster	free	0.111	10	0.050	0.452	128
Hamster	trained, free	0.110	10	0.050	0.455	129
Hamster	quiet, (asleep?)	0.130	12	0.042	0.323	74
Hamster	free	0.092	65	0.061	0.665	93
Mouse	free	0.0198	3 56	0.0245	1.239	93
Mouse		0.0271	5	0.0520	1.919	151
Mouse		0.0120	6	0.0200	1.667	120
*Mouse,	free	0.0105	10	0.0150	1.429	114
Mouse, C57	free	0.0240	7	0.0273	1.138	86
Pony	free	205.	1	19.9	0.097	124
Pony	free	135.	1	23.2	0.172	124
Pony	free	161.	1	45.2	0.281	124

<sup>\*</sup> Excluded as too young
\*\* Excluded as outlier, See text.

Table VI Continued; non-anesthetized animals

Species	Physiological state and details	Weight	Number of animals	volume		Reference
Species	State and details	(kg)	ariiriais	(Littles)	D VVI	neierence
Horse, Morgan	Treadmill trained	476.	5	65.	0.137	150
Horse	standing	422.	2	60.	0.137	131
Horse, Thoroughbred	standing	486.	15	79.	0.142	91
Horse, mixed	standing tied	476.	5	44.	0.100	56
Horse, mixed	Startoning tied	410.	3	77.	0.000	00
Donkey	stanchion	120.	1	19.5	0.163	56
Mule	stanchion	210	1	30.1	0.143	56
Mule	stanchion	210	1	21.7	0.103	56
Rabbit	trained free	3.00	9	1.240	0.413	129
Rabbit	Awake, restrained	3.25	10	1.503	0.463	143
Rabbit	trained, free	2.05	42	1.615	0.787	118, 59
_						
Rat	free	0.400		0.223	0.558	118, 60
Rat	trained, free	0.383		0.215	0.561	129
Rat, white	free	0.310		0.066	0.213	131
Rat	head electrodes	0.300		0.141	0.470	139
Rat	Quiet, awake	0.300		0.153	0.510	138
Rat,	Porton	0.198		0.199	1.008	121
Rat, 18 week	free	0.299		0.225	0.753	117
Rat, 10 week	free	0.211		0.161	0.761	117
Rat, 3-6 mo	plethysmog, whole body	0.303		0.273	0.899	134
Rat, 3-6 mo	plethysmog, whole body	0.303		0.236	0.780	134
Rat	trained, free	0.250		0.140	0.560	60
Rat	trained, free	0.250		0.155	0.625	60
Rat		0.284		0.276	0.971	115
Rat	unrestrained	0.305		0.214	0.701	115
Rat		0.250		0.238	0.950	135
Rat	free	0.198		0.199	1.008	121
Rat	free	0.238		0.388	1.630	135, 73
Rat	free	0.229		0.336	1.467	135, 69
Rat	free	0.111	23	0.228	2.054	135, 69
*Rat, white	free	0.113		0.073	0.646	93
*Rat, 5 week	free	0.052		0.087	1.662	117
*Rat, 7 week	free	0.110	10	0.113	1.028	117
**Cotton rat	free	0.077	27	0.040	0.516	93

<sup>\*</sup> Excluded as too young
\*\* Excluded as outlier, See text.

Table VI Continued; non-anesthetized animals

Species	Physiological state and details	Body Weight (kg)	Number of animals	volume	Minute volume B Wt	Reference
Sheep	stanchion	52.0	1	28.0	0.539	56
Sheep	Thermoneutral water bath	59.9	4	7.10	0.119	53, 100
Sheep	Air	77	1	10.2	0.133	101
Sheep	Air	71	1	9.4	0.133	101
Sheep	Thermoneutral water bath	70	1	11.4	0.163	101
Sheep	20°C environment	52.0	1	28.0	0.538	56
Sheep	Temperature study	29.0	2	13.8	0.477	68
Sheepa	light thiopentone	41.8	13	12.53	0.300	77
Sheep <sup>a</sup>	light thiopentone	41.8	9	11.34	0.271	77
Sheepa	light thiopentone	41.8	2	9.91	0.237	77
Sheep <sup>a</sup>	light thiopentone	41.8	11	11.39	0.272	77
Pig, female, 6 mo	Restrained	170	1	19	0.112	56
Pig, female		225		37	0.164	53, 71
Giraffe	Free standing	544	2	74.3	0.136	141
**Spiny Anteater	♀, Quiet, restrained, asleep?	3.35	2	0.350	0.136	62
**Spiny Anteater	ರೆ, Quiet, restrained, asleep?	3.60	1	0.704	0.196	62
Marine mammals <sup>b</sup>						
Harbour seal	Cheyne-Stokes respiration	27.5		3.97	0.144	104
Porpoise	In captivity	170		9.7	0.057	105
Killer whale	Beached, free	1090	1	47.8	0.044	147

<sup>\*</sup> Excluded as too young

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<sup>\*\*</sup> Excluded as outlier, See text.

<sup>&</sup>lt;sup>a</sup> Body weight value sheep is a minute volume provided as L/min/m<sup>2</sup> body surface and the body weight value is the corresponding weight in kg for one m<sup>2</sup>.

<sup>&</sup>lt;sup>b</sup> Excluded for physiologic reasons, See text.

Table VII

Comparisons of respiratory function and body weight
Summary of data collected; anesthetized animals

		Body	Number		Minute	
_	Physiological	Weight	of	volume		
Species	state and details	(kg)	animals	(Litres)	B Wt	Reference
Cat	pentobarbital Na	3.70	4	0.960	0.259	1
Cat	pentobarbital Na	3.25	6	0.785	0.242	146
Cat	pentobarbital Na	2.80	6	0.410	0.146	130
Cat	pentobarbital Na	2.80	11	0.550	0.196	87
Cat	pentobarbital Na	3.20	9	1.076	0.336	154
Cat	pentobarbital Na	3.40	6	0.840	0.247	88
Cat	pentobarbital Na	3.40	6	0.900	0.265	88
Cat	pentobarbital Na	2.7	5	0.5	0.19	90
Cat	pentobarbital Na	2.5	6	0.468	0.187	156
Cat	ketamine	3.10	10	0.489	0.158	107
Cat	diazepam	3.40	6	1.130	0.332	88
Cat	nembutal	2.55	4	0.250	0.098	152
Cat	chloralose	2.60	4	0.180	0.069	152
Cat	urethane	2.35	4	0.248	0.105	152
Cat	alpha chloralose	2.50	19	0.381	0.152	61
Dog	nembutal	13.5	39	6.22	0.462	57
Dog	nembutal	10.0	8	4.00	0.400	149
Dog	pentobarbital	12.6	4	3.10	0.400	1
Dog, no breed	chloralose/ethyl carbamat		2	1.07	0.115	132
Dog, no breed, females	pentobarbitol	16.8	35	1.81	0.108	97
Dog, no breed, males	pentobarbitol	21.0	42	2.92	0.139	97
Dog, Beagle, 560-700 days	pentobarbital + vetame	9.0	1	1.68	0.186	78
Dog, Beagle, 560-700 days	pentobarbital + vetame	9.0	1	1.76	0.195	78
Dog, Beagle, 560-700 days		9.0	1	1.82	0.202	78
Dog, Beagle, 560-700 days	•	9.0	1	2.16	0.239	78
Dog, Beagle, 560-700 days		9.0	1	2.48	0.276	78
Pig	nembutal	28.	19	6.7	0.239	57
Sheep	NA thiopentone	41.8 <sup>a</sup>	5	6.35	0.152	96
Sheep	Na thiopentone	41.8 <sup>a</sup>	7	6.67	0.159	76
Sheep	Na thiopentone	41.8 <sup>a</sup>	13	12.53	0.300	76
Ferret	pentobarbital	0.314	18	0.195	0.621	70

<sup>&</sup>lt;sup>a</sup> Body weight value for sheep is artificial calculated from a minute volume provided as  $L/min/m^2$  body surface using the formula given by the author for calculating surface area from body weight. The value for minute volume is the quoted value in  $L/min/m^2$  and the body weight is the corresponding weight in kg for one  $m^2$ .

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## **Table VII Continued**

Species	Physiological state and details	Body Weight (kg)	Number of animals	Minute volume (Litres)		Reference
G pig	pentobarbital Na ether	0.690 0.477		0.130 0.154	0.188 0.323	1 93
G pig	ether	0.477	49	0.154	0.323	93
Hamster, male	pentobarbital Na	0.132	27	0.045	0.341	144
Hamster, female	pentobarbital Na	0.136	27	0.040	0.294	144
Hamster	ether	0.096	34	0.048	0.499	144
Hamster	chloralose/ethyl carbamate	0.110	7	0.072	0.650	132
Hamster, 15 week	halothane	0.126	10	0.028	0.221	126
Hamster, 65 week	halothane	0.125	9	0.028	0.225	126
Giraffe	Free standing	544	1	61	0.111	141
Mouse	pentobarbital Na	0.032	0 4	0.0210	0.656	1
Mouse	pentobarbital	0.027	1 5	0.0207	0.764	151
Mouse	chloralose/ethyl carbamate	0.027	0 10	0.0560	2.074	132
Mouse	ether	0.018	8 7	0.0177	0.941	93
Mouse	ether	0.020	7 15	0.0281	1.357	93
Mouse, C57	pentobarbital Na	0.023	0 5	0.0108	0.470	86
Rabbit	pentobarbital Na	2.400	4	0.620	0.258	1
Rabbit	chloralose/ethyl carbamate	1.800	4	0.450	0.250	132
Rabbit	ether	2.069	31	0.800	0.387	93
Rabbit, 0.5-1 yr	pentobarbital Na	4.100	15	0.773	0.188	80
Rabbit, 3-5 yr	pentobarbital Na	4.100	20	0.694	0.169	80
Rat	chloralose/ethyl carbamate		16	0.152	0.707	132
Rat	urethane	0.163	21	0.119	0.732	135
Rat	urethane	0.230	18	0.146	0.635	135
Rat	urethane	0.285	24	0.171	0.600	135
Rat	urethane	0.323	14	0.127	0.393	75, 137
Rat	urethane + allobarbital	0.195	10	0.092	0.472	135, 75, 137
Rat	halothane	0.200	20	0.054	0.270	79
Rat	urethane	0.206	67	0.085	0.416	135, 136
Rat	urethane	0.210	10	0.288	1.371	135
Rat	halothane, light	0.229	10	0.203	0.886	135
Rat	halothane, light	0.220	10	0.132	0.600	135
Rat	halothane, light	0.234	9	0.149	0.637	135
Rat	halothane, light	0.266	8	0.189	0.711	135
Rat	halothane, light	0.280	7	0.115	0.411	135
Rat	pentobarbital Na	0.250	4	0.160	0.640	1

## **Table VII Continued**

Species	Physiological state and details	Body Weight (kg)	Number of animals	volume		Reference
Rat, female, B&W Hooded	pentobarbital Na	0.257	28	0.111	0.432	144
Rat, male, B&W hooded	pentobarbital Na	0.464		0.207	0.446	144
Rat, Fisher, female	urethane	0.233		0.161	0.691	81
Rat, white	ether	0.110		0.076	0.688	93
Rat, white	thiamyl	0.402		0.175	0.435	108
Rat, 102 days	halothane	0.222		0.054	0.243	127
Rat, 538 days	halothane	0.334		0.082	0.246	127
Rat, 815 days	halothane	0.332		0.082	0.247	127
·						
Monkey	sernyl	2.80	2	0.530	0.189	63
Monkey	pentobarbital Na	2.45	4	0.700	0.286	1
Monkey	ketamine	2.61	1	0.480	0.184	84
Monkey	ketamine	3.03	1	0.600	0.198	84
Monkey	ketamine	3.53	1	0.900	0.255	84
Monkey	ketamine	3.33	1	0.900	0.270	84
Monkey	ketamine	3.32	1	0.800	0.241	84
Monkey	ketamine	2.71	1	0.750	0.277	84
Monkey	ketamine	3.66	1	1.100	0.301	84
Monkey	ketamine	3.31	1	1.000	0.302	84
Monkey	ketamine	3.50	1	0.900	0.257	83
Monkey	ketamine	3.50	1	1.000	0.286	83
Monkey	pentobarbital	3.50	1	0.546	0.156	114
Monkey, Rhesus	pentobarbital	7.56	6	1.71	0.226	98,148
Monkey, Rhesus	tileamine	3.30	10	0.462	0.140	64
Monkey, Rhesus	ketamine+acepromazine	3.30	10	0.442	0.134	64
Monkey, Rhesus	ketamine	3.30	10	0.575	0.174	64
Monkey, Rhesus	pentobarbital	4.25	11	1.650	0.388	72, 148
Monkey, Rhesus	pentobarbital	7.60	8	3.860	0.508	148
Monkey, Rhesus	pentobarbital	3.49	4	0.681	0.195	109
Monkey, Rhesus	pentobarbital	4.30	21	0.709	0.165	111
Monkey, Rhesus	phencyclidene	5.80	14	1.791	0.309	116, 148
Monkey, Rhesus	pentobarbital, O <sub>2</sub>	5.05	9	0.900	0.178	119
Monkey, Rhesus	pentobarbital	5.05	9	1.360	0.269	119

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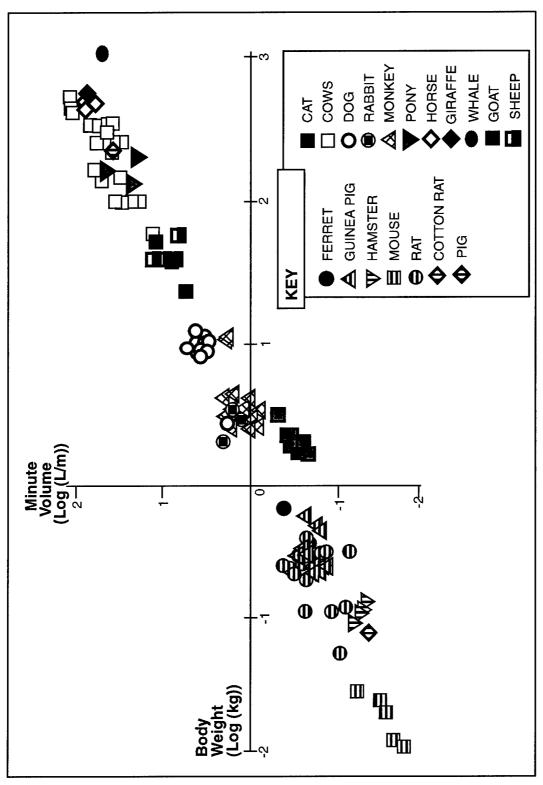


Fig. 1 Body weight and minute volume of representative animal data in the collected data base. The data show an allometric relationship on this Log-Log plot that is linear and flows readily through the data. The data represent a range from a mouse (12 g) to a whale (1090 kg).

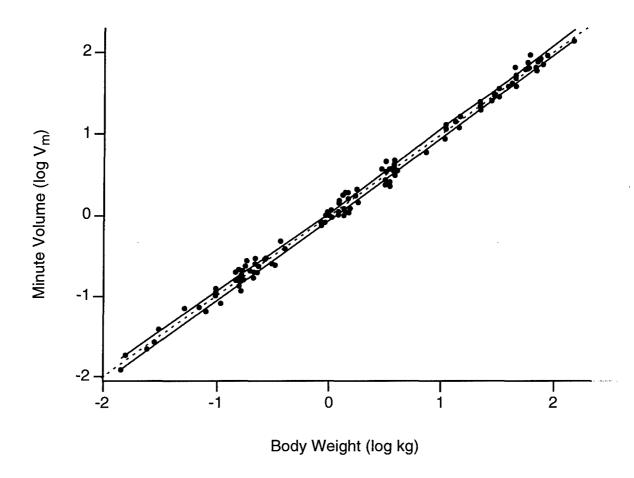


Fig. 2 A statistical plot of the allometric equation for non-anesthetized animals and the statistical error limits. For this plot, the final data set was used. Marine mammals, very young animals and several groups of outlying data have been excluded. The fit of the regression line is very good;  $r^2 = 0.975$ .

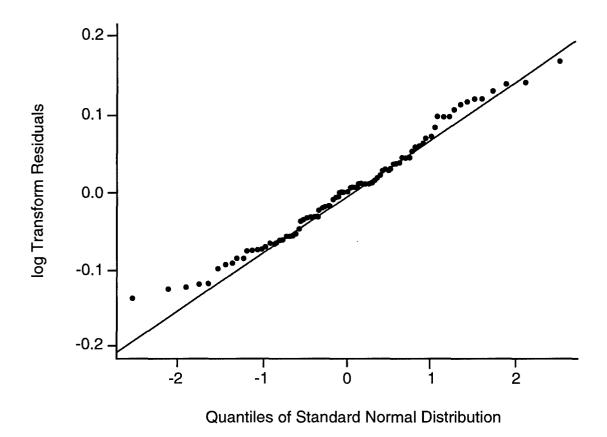


Fig. 3 A quantile plot of the residuals from the fitted line describing the difference between the measured and the predicted logarithmic minute ventilation rates.

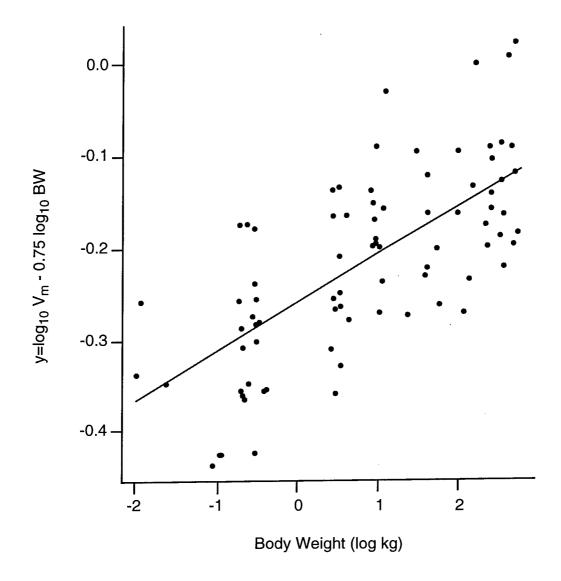


Fig. 4 When the power-law exponent is assumed to be 3/4, the plot of residuals vs body weight indicates an upward trend. This suggests that  $\mathbf{V}_{\mathrm{m}}$  varies with  $\mathbf{B}\mathbf{W}$  with a greater than 3/4 scaling component.

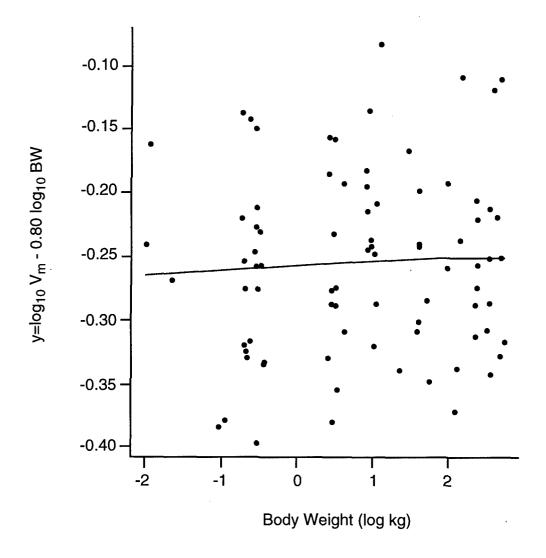


Fig. 5 When the power-law exponent is 0.80, the plot of residuals vs body weight does not show any systematic trends suggesting that all of the variability in the data is accounted for in the allometric equation.

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The relationship between body weight (BW) and respiratory minute volume (V<sub>m</sub>) was reviewed by collecting a data base from the literature of minute volume rates that encompassed species from mice at 12 g body weight to horses and a giraffe at ≈500 kg body weight. The data were separated into anesthetized and non-anesthetized groups and juvenile animals were removed from the non-anesthetized group. The final data set of non-anesthetized animals contained 131 studies representing 2125 animals and 18 species. The data show a power-law (allometric) relationship between the minute volume and body weight. The scaling or allometric parameters in this power-law have been estimated using a linear regression of the logarithms of the minute volume against body weight. The resulting allometric equations were;

$$Log_{10}V_m = -0.286 + 0.802 Log_{10}BW$$
 or  $V_m = 0.518 BW^{0.802}$ 

From these equations a corresponding set of minute volumes were obtained for various body weights of humans eg. 15.6 L/min for a 70 kg human. The results of the analyses were compared to similar studies in the literature. The relationship is recommended for military uses because it is derived from non-anesthetized, young adult mammals which are expected to mimic the soldier.

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